SOLANOPUBAMINE, A STEROIDAL ALKALOID FROM SOLANUM PUBESCENS

G. N. Krishna Kumari, L. Jagan Mohan Rao, K. V. Raja Rao, N. S. Prakasa Rao, K. Kaneko* and H. Mitsuhashi*

Department of Chemistry, Nagarjuna University, Nagarjunanagar 522 510, India; *Faculty of Pharmaceutical Sciences, Hokkaido University, Sapporo 060, Japan)

(Revised received 16 October 1984)

Key Word Index—Solanum pubescens; Solanaceae; steroidal alkaloid; 3β-amino-5α,22αH,25βH-solanidan-23β-ol.

Abstract—Aerial parts of Solanum pubescens yielded a new steroidal alkaloid, solanopubamine, the structure of which was elucidated as 3β -amino- 5α , $22\alpha H$, $25\beta H$ -solanidan- 23β -ol by ¹³C NMR, ¹H NMR, IR, mass spectral analysis and chemical degradation methods.

INTRODUCTION

In our continued chemical examination of the taxon Solanum pubescens Willd. [1-3] we isolated a novel steroidal alkaloid, solanopubamine, from the aerial parts. The alkaloid has the molecular formula $C_{27}H_{46}N_2O$ ([M]⁺, m/z 414) as reported in this paper and its structure has been elucidated as 3β -amino- 5α ,22 αH ,25 βH -solanidan- 23β -ol(3-deoxy- 3β -amino- 5α ,6-dihydroleptinidine, 1).

RESULTS AND DISCUSSION

The mass spectrum of 1 showed diagnostic fragmentations [4, 5] at m/z 166 (100%) and 220 (24%) for a solanidane skeleton with a hydroxyl in either rings E or F. Furthermore, the mass spectral fragments at m/z 370 (5%) and 343 (6%) fixed the hydroxyl on C-23 [6]. The ions at m/z 56 (9%) and 82 (8%) indicated the presence of a 3-amino group [4], accounting for the second nitrogen, one being in the indolizidine moiety.

The ¹H NMR spectrum of 1 at 270 MHz in deuterochloroform plus deuteromethanol showed two tertiary methyl groups at $\delta 0.85$ (3H, s) and 0.89 (3H, s) and two secondary methyl groups at $\delta 0.98$ (3H, d, J=6.5 Hz) and 1.20 (3H, d, J=7 Hz) which were allocated to Me-18, Me-19, Me-21 and Me-27, respectively.

Solanopubamine (1) formed a diacetate, 2 $(C_{31}H_{50}N_2O_3)$. Its 200 MHz ¹H NMR spectrum in deuterochloroform indicated two acetyl signals at δ 1.95 and 2.02. The protons due to the 3N-acetyl group appeared as a broad doublet at δ 5.28 (1H, J=8 Hz) and a singlet at δ 1.95 (3H) and were assigned to NH and NHCOCH₃. The 23 O-acetyl signal appeared at δ 2.02.

The broad low-field signal at δ 3.04 (1H, $W_{1/2} = 18$ Hz) in 1 which shifted on acetylation to δ 3.75 was allocated to the proton on C-3 bearing the amino group [7] and assigned an axial orientation of H-3 from the $W_{1/2}$ values.

The characteristic intramolecularly hydrogen bonded hydroxyl absorption band at $v_{\text{max}}^{\text{KBr}}$ cm⁻¹: 3530 [8] in the IR spectrum of 1 remained unchanged in dilution experiments. The hydroxyl was allocated to C-23, as that was the only available position for a secondary hydroxyl to form

an intramolecular hydrogen bond with the indolizidine nitrogen lone pair. The signal at $\delta 3.83$ (1H, $W_{1/2} = 7$ Hz) in 1 and $\delta 5.02$ in its acetate, 2, were assigned to the proton on C-23 bearing the hydroxyl group. The $W_{1/2}$ value of 7 Hz for H-23 indicated an axial orientation of the OH-23

1 $R = NH_2, R^i = H$

 $2 R = NHAc, R^1 = Ac$

3 $R = OH, R^1 = H$

10

group [6]. The absence of any other signal beyond $\delta 3.00$ except for the protons at C-3 and C-23 indicated that both the protons at C-16 and C-22 must be *trans* to the lone pair of electrons on the nitrogen as in other solanidanes [6].

The presence of the Bohlmann band at $v_{\text{max}}^{\text{KBr}}$ cm⁻¹: 2750 in the IR spectrum of 1 supported the trans-indolizidine fusion of the E and F rings similar to that of solanidine. The signal at $\delta 2.77$ (d, J = 11 Hz) was assigned to the equatorial H-26 by analogy with that of solanogantamine [6].

The 13 C NMR assignment of solanopubamine (1) (Table 1) was based on the data of 3β -amino- 5α -cholestane (4) [9], jurubidine (5) [9], indolizidine (6) [10] and demissidine (7) [11]. The chemical shift values for the A-C ring carbons in 1 were similar to that of 4 and 5 and supported the allocation of the amino group to C-3 and assigns the stereochemistry of rings A-C as all trans. A comparison of the D-F ring signals of 1 with those of 6 and 7 indicated that the observed $\Delta\delta$ values within the reported range [9, 12] are in excellent agreement and supported the axial orientation of OH-23 in 1. The α -(23) and β -(22 and 24) carbons moved downfield by $\Delta\delta$ + 36.4 (δ 65.7), + 4.4 (δ 79.1) and + 2.8 (δ 33.9), respectively. The γ -(20 and 25) carbons showed an upfield shift of $\Delta\delta$ - 5.8 (δ 30.9) and - 3.3 (δ 28.0), respectively.

A study of the effect of the methyl substituent of indolizidine as of cyclohexane was found to be useful in assigning the orientation of the methyl group attached to C-25. These effects were calculated using the $\Delta\delta$ values from cyclohexane to methylcyclohexane [13]. From the

calculated values for 9 and 10, C-25 was expected to resonate at δ 26.2 for axial orientation and at δ 30.7 for equatorial orientation of the Me-27 group.

The chemical shift due to C-25 appeared at δ 28.0 ($\Delta\delta$ – 2.7). The upfield shift from the calculated value can be attributed to the γ -effect experienced by this carbon due to the OH-23 group [12, 13] ($\Delta\delta$ is – 3.3 when compared to the demissidine C-25 signal at δ 31.3). Hence, the orientation of the Me-27 group in 1 was assigned as equatorial. As the expected chemical shift value must be further upfield from the calculated δ 26.2 for an axial methyl group, this possibility was ruled out.

The equatorial orientation was further supported by deamination of 1. Nitrous acid deamination of 1 produced a diol as the major product which was found to be dihydroleptinidine (3) [14] from mp 221-222°, $[\alpha]_D + 29.7^\circ$ (CHCl₃; c 0.39), IR $v_{\text{max}}^{\text{KBr}}$ cm⁻¹: 3600, 3500, 2750, 833 and ¹³C NMR spectral data (Table 1). The formation of 3 supported the orientation of the OH-23 and Me-27 groups as β -axial and α -equatorial, respectively.

Solanopubamine was found to be a stereoisomer of solanogantamine [6, 15] and their non-identity was proved by direct comparison with an authentic sample by mmp and co-TLC. The deamination products from these two compounds were also found to be different (Table 2).

EXPERIMENTAL

All mps are uncorr. ¹H NMR spectra were measured at 270, 100, 99.5 and 199.5 MHz, ¹³C NMR at 25 and 50 MHz.

Table 1. 13C NMR data of compounds 1 and 3-7

Carbon No.	1 (deutero- pyridine)	3 (deutero- chloroform)	4 [9]	5 [9]	6 [10]	7 [11]
1	37.5	37.1	37.8	37.7		37.1
2	32.2	31.6	32.6	32.3	_	31.6
3	50.9	71.4	51.2	50.9		71.3
4	40.1	38.3	39.6	40.1		38.3
5	45.1	45.0	45.7	45.5		45.0
6	28.7	28.7	28.9	28.6		28.8
7	31.5	32.3	32.3	31.7		32.3
8	35.3	35.4	35.6	35.2	_	35.4
9.	54.3	54.5	54.6	54.5	_	54.6
10	35.7	35.6	35.6	35.6	_	35.6
11	21.0	21.1	21.2	21.0		21.1
12	36.9	39.6	40.2 •	37.6		40.2
13	41.4	41.5	42.7	40.6		40.6
14	57.4	57.5	56.4	56.4		57.4
15	27.5	31.6	24.3	30.9	_	33.5
16	69.8	69.6	28.3	80.9	53.9	69.0
17	62.5	62.2	56.6	62.1	20.3	63.3
18	17.1	16.8	12.1	16.5		17.1
19	12.2	12.4	12.4	12.3		12.4
20	30.9	30.7	35.9	42.2	30.1	36.7
21	18.7	18.9	18.7	14.3	_	18.3
22	79.1	79.0	36.3	109.7	64.1	74.7
23	65.7	67.0	23.9	26.0	30.7	29.3
24	33.9	37.1	39.4	25.8	24.2	31.1
25	28.0	26.9	28.1	27.1	25.1	31.3
26	58.7	58.7	22.9	65.1	52.7	60.2
27	21.9	22.4	22.9	16.2		19.5

Compound	Mp	[α] _D	(solvent)	Ref.				
Solanopubamine (1)	263°	+ 30.5°	(methanol)					
Deamination product (3)	221-222°	$+29.7^{\circ}$	(chloroform)					
Dihydroleptinidine	221-224°	+31.3°	(chloroform)	[14]				
Solanogantamine	180°	+35°	(chloroform)	โ6โ				
Deamination product	215-216°	+40.4°	(chloroform)	โ้6ไ				

Table 2. Physical constants of solanopubamine (1) and solanogantamine, their deamination products and dihydroleptinidine

Leaves and stems of *S. pubescens* Willd. were collected at Nagarjuna Sagar in Andhra Pradesh in Feb. 1982. Air dried, powdered material (5 kg) was extracted with *n*-hexane and MeOH successively.

The concd MeOH extract was separated into phenolic and non-phenolic parts with neutral lead acetate. The non-phenolic part, after de-leading, was refluxed with 10% HCl at 100° for 2 hr, cooled, basified with 10% NaOH soln to pH 8, allowed to stand at room temp., filtered and washed with H2O. The brown residue was chromatographed on a column of silica gel using C₆H₆, $C_6H_6-Me_2CO$ as solvents. The benzene-Me₂CO (1 + 1) fraction yielded 1. It was recrystallized from Me₂CO-MeOH to yield white needles, mp 263°. (Found: C, 78.3; H, 11.0; N, 6.60. $C_{27}H_{46}N_2O$ requires: C, 78.2; H, 11.8 and N, 6.7%.) $[\alpha]_D + 30.5^\circ$ (MeOH); IR $v_{\text{max}}^{\text{KBr}}$ cm⁻¹: 3530 (m), 3075 (m), 2950 (s), 2900 (s), 2850 (m), 2820 (m), 2750 (m), 1530 (m), 1460 (m), 1400 (s), 1330 (s), 1240 (w), 1170 (w), 1150 (w), 1030 (m), 835 (m) and 825 (m); ¹H NMR (270 MHz, CDCl₃ + CD₃OD): δ 0.85 (3H, s), 0.89 (3H, s), 0.98 (3H, d, J = 6.5 Hz); 1.20 (3H, d, J = 7 Hz), 2.77 (1H, d, J= 11 Hz), 2.86 (1H, $W_{1/2}$ = 18 Hz), 3.04 (1H, $W_{1/2}$ = 18 Hz), 3.83 (1H, $W_{1/2} = 7$ Hz); MS m/z (rel. int.): 414 [M]⁺ (27.1), 413 (7.1), 399 (4.9), 396 (1.8), 370 (4.8), 343 (5.5), 220 (24), 166 (100), 82 (8.3),

Acetylation of 1. To a soln of 40 mg 1 in 1 ml C_5H_5N , 0.5 ml Ac_2O was added and kept at room temp. for 40 hr. Excess reagent was removed under red. pres. and recrystallized from MeOH to yield 40 mg white needles, mp 232°. (Found: C, 74.72; H, 9.97; N, 5.56. $C_{31}H_{50}N_2O_3$ requires: C, 74.69; H, 10.00; N, 5.60%.) IR $\nu_{\text{max}}^{\text{CHCl}_3}$ cm⁻¹: 3200, 1720, 1680, 1620, 1240; ¹H NMR (200 MHz, CDCl₃): δ 0.79 (3H, s), 0.89 (3H, s); 0.93 (3H, d, J = 6.3 Hz), 1.13 (3H, d, J = 6.8 Hz), 1.95 (3H, s), 2.02 (3H, s), 2.65 (1H, $W_{1/2}$ = 18 Hz); 2.68 (1H, d, J = 11 Hz), 3.75 (1H, $W_{1/2}$ = 20 Hz), 5.02 (1H, $W_{1/2}$ = 8 Hz). 5.28 (1H, d, J = 8 Hz).

Nitrous acid deamination of 1. To a soln of 20 mg 1 in 2 ml 50% HOAc, 200 mg NaNO₂ was added and kept at room temp. for 18 hr. The reaction mixture was diluted with H_2O , basified with NH₃ and extracted with CHCl₃. The CHCl₃ extract was washed

with H₂O and dried over Na₂SO₄. The major product 3, was separated by prep. TLC using C_6H_6 – Me_2 CO (9:1) (R_f 0.32) and recrystallized from Me₂CO to yield 10 mg white plates, mp 221–222°; $[\alpha]_D^{2D}$ + 29.7° (CHCl₃; c 0.39); IR ν_{max}^{KBr} cm⁻¹: 3600, 3500, 2750, 833.

REFERENCES

- Krishna Kumari, G. N., Jagan Mohan Rao, L. and Prakasa Rao, N. S. (1984) Phytochemistry 23, 2701.
- Krishna Kumari, G. N., Jagan Mohan Rao, L. and Prakasa Rao, N. S., Phytochemistry (in press).
- 3. Krishna Kumari, G. N., Jagan Mohan Rao, L. and Prakasa Rao, N. S., J. Nat. Prod. (in press).
- Budzikiewicz, H., Djerassi, C. and Williams, D. H. (1964) Structure Elucidation of Natural Products by Mass Spectrometry Vol. 2, p. 5. Holden-Day, San Francisco.
- Pakrashi, S. C., Chakravarty, A. K. and Ali, E. (1977) Tetrahedron Letters 645.
- Pakrashi, S. C., Chakravarty, A. K., Ali, E., Dhar, T. K. and Dan, S. (1978) J. Indian Chem. Soc. 15, 1109.
- Bird, G. J., Collins, D. J., Eastwood, F. W. and Swan, J. M. (1978) Tetrahedron Letters 159.
- 8. Schreiber, K. and Ripperger, H. (1967) Chem. Ber. 100, 1381.
- Bird, G. J., Collins, D. J., Eastwood, F. W. and Exner, R. H. (1979) Aust. J. Chem. 32, 797.
- Wenkert, E., Brinda, J. S., Chang, C. J., Cochron, D. W. and Schell, F. W. (1974) Acc. Chem. Res. 7, 43.
- Radeglia, R., Adam, G. and Ripperger, H. (1977) Tetrahedron Letters 903.
- Eggert, H., Van Antwerp, C. L., Bhacca, N. S. and Djerassi, C. (1976) J. Org. Chem. 41, 71.
- Stothers, J. B. (1972) Carbon-13 NMR Spectroscopy. Academic Press, New York.
- 14. Ripperger, H. and Schreiber, K. (1969) Chem. Ber. 102, 4080.
- Chakravarty, A. K., Das, B., Ali, E. and Pakrashi, S. C. (1984)
 J. Chem. Soc. Perkin Trans. 1 3, 467.